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Dispiro-1,2,4,5-tetraoxane Analogues of WR 148999.
7,8,15,16-Tetraoxadispiro[5.2.5.2]hexadecanes Substituted at the 1 and 10 Positions with Unsaturated and Polar Functional Groups

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Synthesis and Antimalarial Activity of 11 Dispiro-1,2,4,5-tetraoxane Analogues of WR 148999. 7,8,15,16-Tetraoxadispiro[5.2.5.2] hexadecanes Substituted at the 1 and 10 Positions with Unsaturated and Polar Functional Groups

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Eleven novel dispiro-1,2,4,5-tetraoxanes 3 bearing unsaturated and polar functional groups were designed to enhance the oral antimalarial activity of the prototype tetraoxane 2 (WR 148999). With the exception of 3g and 3h, tetraoxanes 3 were available via the peroxidation of corresponding cyclohexanone derivatives in H₂SO₄/CH₃CN. Tetraoxanes 3g and 3h were prepared by hydrolysis of ester tetraoxanes **3e** and **3i**, respectively. Five of the 11 tetraoxanes were inactive, but six tetraoxanes had IC₅₀ values of 6-26 nM against the K1 and NF54 strains of Plasmodium falciparum compared to corresponding IC₅₀ values of 28 and 39 nM for 2, and 10 and 12 nM for artemisinin (1). Ester tetraoxane 3e was the most active in vitro, some 2-fold more potent than 1. However, none of the six tetraoxanes active in vitro were as effective as either 1 or 2 in vivo; at single doses of 100 mg/kg most possessed little to no vivo activity in mice infected with *Plasmodium berghei*. Unsaturated tetraoxane 3a was uniquely more active when administered per os (po) than subcutan (sc). For this series of tetraoxanes, the discrepancy between vitro and vivo activities underscores the limitations of conclusions drawn solely from in vitro antimalarial data and illustrates a practical benefit of complementary single-dose in vivo antimalarial screens.

The discovery of artemisinin (qinghaosu, 1), a naturally occurring endoperoxide sesquiterpene lactone, stimulated a substantial effort to elucidate its structureactivity relationship (SAR) and initiated an ongoing quest to identify structurally simple and synthetically accessible antimalarial peroxides. Among synthetic antimalarial peroxides, dispiro-1,2,4,5-tetraoxanes such as 2 (WR 148999) are notable in that they differ considerably in structure from 1, are readily prepared in one step from substituted cyclohexanones, and possess antimalarial activity comparable to 1.2,3

The poor to modest oral bioavailability⁴ of artemisinin and its semisynthetic derivatives has been attributed to a high first-pass drug metabolism. Whether the poor oral antimalarial activity of tetraoxane 2 is due to poor absorption or rapid inactivating metabolism is unknown. However, the excellent ip activity of 2³ suggests that this tetraoxane has some resistance to inactivating first-pass hepatic drug metabolism as a major exit

mechanism for drugs from the peritoneal cavity is by way of the portal circulation.⁵ It follows that the low oral activity of 2 may be due either to inactivating gut metabolism or to an inadequate dissolution rate which is a function of its intrinsic solubility, stability to stomach acid, and formulation.

Design

To address the hypothesis that tetraoxane oral bioavailability is a function of oral absorption, we envisioned the synthesis of more water soluble tetraoxanes bearing polar functional groups. 6 These analogues might be more easily absorbed than their alkyl-substituted counterparts, an important consideration if lack of absorption is a significant factor in the low oral activity of tetraoxane 2. Using tetraoxane 2 as a prototype, we designed target dispiro-1,2,4,5-tetraoxanes (7,8,15,16tetraoxadispiro[5.2.5.2]hexadecanes) 3 bearing unsatur-

ated and functional groups at the 1 and 10 positions. As exemplified by 2, dispiro-1,2,4,5-tetraoxanes derived from 2-alkyl cyclohexanones are always formed as single centrosymmetric stereoisomers.^{2,7} We anticipated that this predictable stereochemical outcome would hold true for tetraoxanes 3 as well.

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Table 1. Antimalarial Activity against P. falciparum in Vitro

		IC ₅₀ ,	nM ^a
cmpd	R	K1	NF54
3a	CH=CH ₂	19	23
3b	C≡CH	13	13
3c	C_6H_5	>200	>200
3d	CH_2COOEt	14	16
3e	COOEt	6.2	6.5
3f	CH_2COOH	>200	>200
3g	COOH	>200	>200
3h	OH	>200	>200
3 i	$OCOC_6H_5$	>200	>200
3j	OMe	15	16
3k	$OCH_2C_6H_5$	18	26
2	Н	28	39
1		10	12

^a Average of n = 2.

Tetraoxanes 3 were designed as derivatives of 2 by replacing one methyl hydrogen atom of each of the two methyl groups in 2 with unsaturated (3a-3c), carboxylic (3d-3g, 3i), hydroxylic (3h) and ether (3j, 3k) substituents (Table 1). Five of the target tetraoxanes could also conceivably function as prodrugs in vivo. For example, 3j and 3k could both be converted to 3h via metabolic O-dealkylation reactions. In addition, 3d, 3e, and 3i could be converted to 3f, 3g, and 3h, respectively, via plasma esterase enzymes.

Chemistry

Nine tetraoxanes were prepared according to the peroxidation method developed by McCullough et al.⁸ in which cyclohexanones are treated with hydrogen peroxide in H₂SO₄/CH₃CN. The unoptimized yields ranged from 4 to 12%. Tetraoxanes **3g** and **3h** were not formed using this method and were instead indirectly prepared by the hydrolysis of tetraoxane esters **3e** and **3i** in yields of 72 and 60%, respectively. The failure to form **3g** and **3h** via acid-catalyzed peroxidation is possibly associated with formation of persistent perhydrates, in agreement with reported data⁹ in which 2-chloro- and 2-bromocyclohexanone afforded isolable perhydrates instead of tetraoxanes due to the formation of an intramolecular hydrogen bond between the hydroperoxide hydrogen and the halogen substituent.

Tetraoxanes **3a**–**3k** are white crystalline solids stable at room temperature for months if not years. In ¹H NMR spectra of these tetraoxanes, as is also the case for tetraoxane **2**,² a characteristic peak for the two methine protons at the 1 and 10 positions falls between 2.7 and 3.4 ppm. In the corresponding ¹³C NMR spectra, a diagnostic signal is present at 108–110 ppm, corresponding to the spiro quaternary carbon atoms.^{2,7}

Antimalarial Activity

The in vitro antimalarial data (Table 1) reveal that six of the tetraoxanes were at least as potent as the parent tetraoxane 2. Of these, 3b, 3d, and 3j were nearly as potent as artemisinin, and 3e was notably more potent than artemisinin. From these data, we can conclude that the presence of alkene (3a), alkyne (3b), and ether (3j, 3k) moieties maintain or enhance antimalarial potency whereas carboxylic acid (3f, 3g) and alcohol (3h) moieties decrease antimalarial potency. The situation for esters is less clear; ethyl esters 3d and 3e had good activity, whereas 3i, the dibenzoate ester of

Table 2. Antimalarial Activity against P. berghei in Vivo

cmpd^a	activity (%) ^b		survival (days) ^c	
	sc	po	sc	po
3a	15	93		7
3b	9	0		
3 d	0	0		
3 e	0	0		
3j	100	18	24.7	
3j 3k	0	0		
2	100	99.7	27.7	7.7
1	100	98	14	7.3

^a Compounds in a solution or a suspension containing 3% ethanol and 7% Tween 80 were administered on day 1 post-infection at a single dose of 100 mg/kg. ^b Reduction of parasitemia on day 3 post-infection. ^c On average, mice in a control group survive 5.2 days post-infection.

3h, was inactive. We also note that the six active tetraoxanes, like artemisinin, were more potent against the chloroquine-resistant K1 strain than against the drug-sensitive NF54 strain of *Plasmodium falciparum*, although in each case, the potency difference was small.

The six active tetraoxanes, along with 1 and 2 as controls, were selected for in vivo antimalarial evaluation in mice infected with the ANKA strain of Plasmodium berghei. Consistent with existing multiple-dose Thompson test data,^{2,3} the vivo antimalarial efficacies for 1 and 2 differed only slightly. From the vivo data (Table 2), it is clear that none of these functionalized tetraoxanes possessed useful oral antimalarial activity and all were less effective than either 1 or 2. We attribute the complete lack of vivo activity for ester tetraoxanes **3d** and **3e** to in vivo chemical or enzymatic hydrolysis to the inactive tetraoxane acids 3f and 3g, respectively. Although methyl ether tetraoxane 3j had good vivo activity sc, it was almost completely inactive po due probably to first-pass O-dealkylative metabolism to the inactive tetraoxane alcohol 3h. Unsaturated tetraoxane **3a** was uniquely more active po than sc, due possibly to formation of some active metabolite.

Discussion

In this work, we identified six functionalized tetraoxanes with in vitro potency equal or superior to prototype tetraoxane 2 and one diester tetraoxane (3e) with in vitro potency superior to 1. It is conceivable that some of the tetraoxanes were well absorbed orally but suffered from a lack of metabolic stability. Nevertheless, the poor vivo antimalarial activity of these tetraoxanes provides no justification for additional experiments (i.e., blood level determinations) to assess their oral bioavailability. For this series of tetraoxanes, the discrepancy between vitro and vivo activities underscores the limitations of conclusions drawn solely from in vitro antimalarial data and illustrates how the single-dose in vivo antimalarial screen used in this study provided a necessary and more rigorous test of antimalarial potential.

As these functionalized tetraoxanes were based on tetraoxane **2** as a structural template, they manifest the advantage of both defined and predictable stereochemistry but also the liability of their centrosymmetric symmetry in which each tetraoxane (ketone diperoxide) necessarily bears two functional groups present singly in the starting material cyclohexanones. Existing tetraoxane synthetic methods¹⁰ do not allow for the synthesis of nonsymmetrical tetraoxanes. Even with this

limitation, incorporation of unsaturated and polar functional groups at other positions in dispiro-1,2,4,5tetraoxanes is possible, and this work provides a guide to functional group selection in the search for more orally active tetraoxane antimalarials.

Experimental Section

Melting points are uncorrected. Unless noted otherwise, ¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded on a Varian XL-300 spectrometer using CDCl₃ as a solvent. All chemical shifts are reported in parts per million (ppm) and are relative to internal (CH₃)₄Si for ¹H and CDCl₃ (77.0 ppm) for ¹³C NMR. Microanalyses were performed by M-H-Wlaboratories, Phoenix, AZ. 2-Allyl- and 2-carboethoxymethylcyclohexanone were purcased from Aldrich Chemical Co. 2-Benzyl-, 2-propargyl-, 2-carboethoxyethyl-, and 2-benzyloxymethylcyclohexanone were prepared by enamine alkylation reactions described by Stork et al.11 2-Hydroxymethylcyclohexanone was prepared from cyclohexanone and paraformaldehyde. 12 2-Benzoyloxymethylcyclohexanone was synthesized by acylation of 2-hydroxymethylcyclohexanone with benzoyl chloride in pyridine/dichloromethane. 2-Carboxyethyland 2-carboxymethylcyclohexanone were prepared by hydrolysis of the corresponding commercially available ethyl esters in KOH/MeOH followed by acidification using concentrated HCl.

General Procedure⁸ for the Preparation of Tetraoxanes 3a-3f and 3i-3k. A cold (-20 °C) solution of a parent cyclohexanone (10 mmol) in acetonitrile (2 mL) was added dropwise to a stirred, cold (-30 °C) solution of 50% hydrogen peroxide (0.60 mL, 11 mmol) and concentrated sulfuric acid (1 mL) in acetonitrile (4 mL). After being stirred for an additional 1 h at -30 to -20 °C, the solution was kept at -20°C for 2 days. If the required product precipitated or crystallized out of the reaction solvent (3a, 3c, 3e, 3f, 3i, 3k), a simple filtration and recrystallization sufficed for purification. If the product tetraoxane did not precipitate (3b, 3d, 3j), extraction with dichloromethane and subsequent silica gel flash column chromatography eluting with petroleum ether/ether (9:1) were applied to isolate the product. In each case, product tetraoxanes were isolated as white crystalline solids.

- 1,10-Bis(2-propenyl)-7,8,15,16-tetraoxadispiro[5.2.5.2]hexadecane (3a): yield 5%; mp 101-102 °C (CH₃CN); ¹H NMR 1.05-1.45 (m, 4H), 1.46-1.82 (m, 12H), 1.85-2.15 (m, 2H), 2.40-2.70 (m, 2H), 2.80-3.10 (m, 2H), 4.90-5.17 (m, 4H), 5.55-5.90 (m, 2H); ¹³C NMR 22.31, 24.54, 27.01, 29.91, 32.06, 43.27, 109.28, 116.46, 136.84. Anal. (C₁₈H₂₈O₄) C, H.
- 1,10-Bis(2-propynyl)-7,8,15,16-tetraoxadispiro[5.2.5.2]hexadecane (3b): yield 10%; mp 167-168 °C dec (CH₃CN); ¹H NMR 1.15-2.40 (m, 20H), 2.50-2.79 (m, 2H), 2.80-3.04 (bs, 2H); ¹³C NMR 17.54, 22.18, 24.59, 27.19, 29.83, 42.97, 69.44, 82.51, 108.67. Anal. (C₁₈H₂₄O₄) C, H.
- 1,10-Dibenzyl-7,8,15,16-tetraoxadispiro[5.2.5.2]hexa**decane (3c):** yield 13%; mp 182-184 °C dec (CH₃CN); ¹H NMR 1.10-1.42 (m, 4H), 1.48-1.78 (m, 10H), 1.82-1.98 (m, 2H), 2.34-2.53 (m, 2H), 3.03-3.19 (m, 2H), 3.20-3.37 (m, 2H), 7.09-7.37 (m, 10H); ¹³C NMR 22.36, 24.61, 26.66, 30.08, 33.71, 45.65, 109.31, 125.99, 128.29, 129.21, 140.24. Anal. (C₂₆H₃₂O₄)
- 1,10-Bis(carboethoxyethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3d): yield 9%; mp 87 °C; ¹H NMR 1.26 (t, J = 7.2 Hz, 6H), 1.20–1.85 (m, 18H), 1.90–2.50 (m, 6H), 2.80-3.15 (bs, 2H), 4.13 (q, J = 7.1 Hz, 4H); 13 C NMR 14.17, 23.08, 24.39, 27.31, 29.64, 29.72, 32.59, 42.91, 60.24, 109.55, 173.40. Anal. (C₂₂H₃₆O₈) C, H.
- 1,10-Bis(carboethoxymethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3e): yield 4%; mp 110 °C (pentane); ¹H NMR 1.26 (t, J = 7.1 Hz, 6H), 1.20–1.85 (m, 14H), 2.05– 2.40 (m, 4H), 2.70-2.89 (m, 2H), 2.90-3.12 (bs, 2H), 4.13 (q, J = 7.1 Hz, 4H); ¹³C NMR 14.18, 22.17, 24.70, 28.36, 29.89, 33.34, 40.33, 60.50, 108.80, 172.38. Anal. $(C_{20}H_{32}O_8)$ C, H.
- 1,10-Bis(carboxyethyl)-7,8,15,16-tetraoxadispiro[5.2.5.2]**hexadecane (3f):** yield 12%; mp 178 °C (CH₃OH:H₂O, 1:1);

¹H NMR (DMSO-*d*₆) 1.05–1.80 (m, 18H), 1.81–2.05 (bs, 2H), 2.06-2.40 (m, 4H), 2.70-2.95 (bs, 2H); ¹³C NMR (DMSO-d₆) $22.19,\, 23.00,\, 24.25,\, 27.16,\, 29.64,\, 32.06,\, 42.58,\, 109.51,\, 174.89.$ Anal. $(C_{18}H_{28}O_8)$ C, H.

1,10-Bis(benzoyloxymethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3i): yield 10%; mp 195 °C dec (ether: pentane, 1:1); ¹H NMR 1.15–2.01 (m, 14H), 2.04–2.32 (bs, 2H), 2.90-3.25 (bs, 2H), 4.10-4.49 (bs, 2H), 4.50-4.80 (bs, 2H), 7.30-7.75 (m, 6H), 7.90-8.30 (m, 4H); ¹³C NMR 22.03, 24.55, 25.72, 30.04, 43.30, 62.90, 109.13, 128.32, 129.56, 130.10, 132.92, 166.42. Anal. (C₂₈H₃₂O₈) C, H.

1,10-Bis(methoxymethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3j): yield 8%; mp 117–118 °C; ¹H NMR 1.10-2.05 (m, 16H), 2.75-3.00 (bs, 2H), 3.32 (s, 6H), 3.15-3.40 (m, 2H), 3.55-3.80 (m, 2H); ¹³C NMR 22.17, 24.38, 25.84, 29.90, 43.91, 58.84, 70.36, 109.14. Anal. (C₁₆H₂₈O₆) C, H.

1,10-Bis(benzyloxymethyl)-7,8,15,16-tetraoxadispiro-**[5.2.5.2]hexadecane (3k):** yield 10%; mp 131 °C (pentane); ¹H NMR 1.20–1.82 (m, 12H), 1.85–2.10 (bs, 4H), 2.70–3.00 (bs, 2H), 3.20-3.55 (bs, 2H), 3.60-3.95 (bs, 2H), 4.30-4.70 (m, 4H), 7.15-7.50 (m, 10H); ¹³C NMR 22.17, 24.39, 26.02, 29.93, 44.06, 68.22, 73.14, 109.18, 127.52, 127.54, 128.32, 138.35. Anal. $(C_{28}H_{36}O_6)$ C, H.

Procedure for Preparation of 3g and 3h. A mixture of diester tetraoxane 3e or 3i (1 mmol), dichloromethane (1 mL), MeOH (10 mL), and KOH (6 mmol, 0.35 g) dissolved in water (1 mL) was heated to 70 °C for 1 h. Diacid tetraoxane 3g was obtained by adding concentrated HCl (1 mL) to the cooled reaction mixture followed by filtration and recrystallization from DMSO. Tetraoxane diol 3h was obtained by adding water (20 mL) to the cooled reaction mixture followed by filtration and recrystallization from MeOH. Tetraoxanes 3g and 3h were isolated as white crystalline solids.

1,10-Bis(carboxymethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3g): yield 72%; mp 198–199 °C dec; ¹H NMR (DMSO-d₆) 1.10–1.80 (m, 14H), 1.90–2.25 (m, 4H), 2.00-2.20 (m, 2H), 2.75-3.00 (bs, 2H); ¹³C NMR (DMSO- d_6 , $30\ ^{\circ}\text{C)}\ 21.72,\ 23.89,\ 27.96,\ 29.07,\ 32.79,\ 39.68,\ 108.44,\ 172.88.$ Anal. $(C_{16}H_{24}O_8)$ C, H.

1,10-Bis(hydroxymethyl)-7,8,15,16-tetraoxadispiro-[5.2.5.2]hexadecane (3h): yield 60%; mp 182 °C dec; ¹H NMR (DMSO- d_6) 1.05-2.10 (m, 16H), 2.56-2.89 (bs, 2H), 3.05-3.32 (m, 2H), 3.60-3.89 (m, 2H), 4.54 (t, J = 5.3 Hz, 2H); ¹³C NMR (DMSO-*d*₆) 21.93, 23.62, 25.13, 29.19, 45.85, 58.34, 108.99. Anal. C₁₄H₂₄O₆: C, H.

Antimalarial Screens. In vitro antimalarial activity against the drug-sensitive NF54 and chloroquine-resistant K1 strains of P. falciparum was determined by a modified tritiated hypoxanthine uptake method of Desjardins et al.¹³ as previously described in detail.¹⁴ Data were expressed as IC₅₀ values, estimated as described by Huber and Koella.15 In vivo antimalarial activity was assessed using Moro SPF mice infected with the ANKA strain of *P. berghei* as described by Ridley et al.¹⁴ Groups of three mice were treated on day 1 post-infection with tetraoxanes dissolved or suspended in 3% ethanol and 7% Tween 80. Tetraoxanes were administered as single 100 mg/kg doses sc and po. Antimalarial activity was measured by percent reduction in parasitemia on day 3 post-infection and by survival times compared to an untreated control group.

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References

(1) For recent reviews, see: (a) Jefford, C. W. Peroxidic Antimalarials. Adv. Drug Res. 1997, 29, 271–325. (b) Casteel, D. A. Antimalarial Agents. In *Burger's Medicinal Chemistry and Drug Discovery;* Wolff, M. E., Ed.; John Wiley & Sons: New York, 1997; Vol. 5, pp 3–91. (c) Cumming, J. N.; Ploypradith, P.;

- Chem. 1992, 35, 3023-3027.
- Vennerstrom, J. L. Unpublished results.
- (a) Titulaer, H. A. C.; Zuidema, J.; Lugt, C. B. Formulation and Pharmacokinetics of Artemisinin and its Derivatives. *Int. J.* Pharmaceut. 1991, 69, 83–92. (b) White, N. J. Clinical Pharmacokinetics and Pharmacodynamics of Artemisinin and Derivatives. *Trans. R. Soc. Trop. Med. Hyg.* **1994**, *88*, 41–43. (c) Li, Q.-G.; Peggins, J. O.; Fleckenstein, L. L.; Masonic, K.; Heiffer, M. H.; Brewer, T. G. The Pharmacokinetics and Bioavailability of Dihydroartemisinin, Arteether, Artemether, Artesunic Acid and Artelinic Acid in Rats. *J. Pharm. Pharmacol.* **1998**, *50*, 173–
- Gibaldi, M. Biopharmaceutics and Clinical Pharmacokinetics, 4th ed.; Lea and Febriger: Philadelphia, 1991; pp 86-87.
- Although reports of functionalized tetraoxanes are rare in the literature, tetraoxanes containing carboxylic acid, ester, and ketone functional groups are known. (a) Fichter, F.; Lurie, S. Lävulinsäure-äthylester-keton-peroxid. *Helv. Chim. Acta* **1931**, 14, 1445–1448. (b) Pummerer, R.; Ebermayer, G.; Gerlach, K. Über das Lävulinsäure-peroxid aus Kautschuk (XII. Mitteil.). *Chem. Ber.* **1931**, *64*, 804–809. (c) White, H. M.; Colomb, H. O., Jr.; Bailey, P. S. Ozonation of 2,5-Diphenylfuran. J. Org. Chem. **1965**, 30, 481–486. (d) House, H. O.; Lee, J. H. C.; VanDerveer, D.; Wissinger, J. E. Perhydroazulenes. 5. Preparation of Perhy-
- droazul-9(10)-en-4-one. *J. Org. Chem.* **1983**, *48*, 5285–5288. (a) Bladon, P.; McCullough, K. J.; Morgan, A. R.; Nonhebel, D. C.; Pauson, P. L.; White, G. J. Ketone-derived Peroxides. Part

- IV. Structural Studies of Cyclic Di- and Tri-peroxides Derived from Ketones. *J. Chem. Res. (M)* **1980**, 3701–3716. (b) McCullough, K. J. Unpublished X-ray crystallographic data for **2**.
- McCullough, K. J.; Morgen, A. R.; Nonhebel, D. C.; Pauson, P. L.; White, G. J. Ketone-derived peroxides. Part I. Synthetic Methods. J. Chem. Res. (M), 1980, 0601-0628
- (a) Ganeshpure, P. A.; Adam, W. \(\alpha \)-Hydroxy Hydroperoxides (Perhydrates) as Oxygen Transfer Agents in Organic Synthesis. Synthesis 1996, 179-188. (b) Kharasch, M. S.; Sosnovsky, G. Structure of Peroxides Derived from Cyclohexanone and Hydrogen Peroxide. J. Org. Chem. 1958, 23, 1322–1326.
- (10) Dong, Y.; Vennerstrom, J. L. Dispiro-1,2,4,5-Tetraoxanes via Ozonolysis of Cycloalkanone O-Methyl Oximes: A Comparison with the Peroxidation of Cycloalkanones in Acetonitrile-Sulfuric Acid Media. J. Org. Chem. 1998, 63, 8582-8585.
- (11) Stork, G.; Brizzolara, A.; Landesman, H.; Szmuszkovicz, J.; Terrell, R. The Enamine Alkylation and Acylation of Carbonyl Compounds. *J. Am. Chem. Soc.* **1963**, *85*, 207–221. Findlay, J. A.; Desai, D. N.; Macaulay, J. B. Thermally induced
- crossed aldol condensations. Can. J. Chem. 1981, 59, 3303-3304.
- Desjardins, R. E.; Canfield, C. J.; Haynes, J. D.; Chulay, J. D. Quantitative assessment of antimalarial activity in vitro by a semiautomated microdilution technique. Antimicrob. Agents Chemother. 1979, 16, 710-718.
- (14) Ridley, R. G.; Matile, H.; Jaquet, C.; Dorn, A.; Hofheinz, W.; Leupin, W.; Masciadri, R.; Theil, F.-P. Richter, W. F.; Girometta, M.-A.; Guenzi, A.; Urwyler, H.; Gocke, E.; Potthast, J.-M.; Csato, M.; Thomas, A.; Peters, W. Antimalarial Activity of the Bisquinoline $trans-N^1,N^2$ -Bis(7-Chloroquinolin-4-yl)Cyclohexane-1,2-Diamine: Comparison of Two Stereoisomers and Detailed Evaluation of the S,S Enantiomer, Ro 47-7737. Antimicrob. Agents Chemother. **1997**, *41*, 677–686. (15) Huber, W.; Koella, J. C. A comparison of three methods of
- estimating EC_{50} in studies of drug resistance in malaria parasites. *Acta Trop.* **1993**, *55*, 257–261.

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